

Deficit irrigation and ascorbic acid in guava cultivation at different development stages

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ABSTRACT: In the semi-arid region of Northeast Brazil, irregular rainfall throughout the year has become a challenge for agricultural production. Given the above, this study evaluated the effects of foliar application of ascorbic acid on the morphophysiology, production components and fruit quality of guava cv. Paluma under irrigation strategies with water deficit. Treatments consisted of a combination of seven irrigation strategies with water deficit - IMS (SE - plants under full irrigation throughout the cycle - 100% crop evapotranspiration - ETC; VE - plants irrigated with water deficit (50% ETC) in the vegetative stage; FL - flowering stage; FR - fruiting stage; MAT - in the fruit maturation stage; VE/FL - in the vegetative and flowering stages; VE/FR - vegetative and fruiting stages) and four concentrations of ascorbic acid - AsA (0, 200, 400 and 600 mg L⁻¹), distributed in a randomized block design, in a split-plot scheme with three replicates, with each plot consisting of 3 usable plants. Water deficit in the vegetative/fruiting stage negatively affected growth in stem diameter, crown volume, crown diameter and vegetative vigor index, average fruit weight, and polar and equatorial diameter of guava fruits at 390 days after transplanting. Foliar application of ascorbic acid at a concentration of 600 mg L⁻¹ promoted positive effects on total fruit weight and yield of guava at 390 days after transplanting. Water deficit in the maturation stage stimulated the synthesis of vitamin C and soluble solids in guava plants at 390 days after transplanting.

Key words: *Psidium guajava* L., non-enzymatic compound, water deficit.

Irrigação deficitária e ácido ascórbico no cultivo de goiabeira em diferentes estádios de desenvolvimento

RESUMO: No semiárido do Nordeste brasileiro a irregularidade das chuvas ao longo do ano tem se tornado um desafio para a produção agrícola. Diante do exposto, objetivou-se avaliar os efeitos da aplicação foliar de ácido ascórbico na morfofisiologia, componentes de produção e qualidade dos frutos de goiabeira cv. Paluma sob estratégias de irrigação com déficit hídrico. Os tratamentos foram constituídos da combinação de sete estratégias de irrigação com déficit hídrico - EMI (SE - plantas sob irrigação plena durante todo o ciclo - 100% da evapotranspiração da cultura - ETC; VE - plantas irrigadas com déficit hídrico (50% da ETC) na fase vegetativa; FL - floração; FR - frutificação; MAT - na fase maturação dos frutos; VE/FL - nas fases vegetativa e de floração; VE/FR - vegetativa e frutificação) e quatro concentrações de ácido ascórbico - AsA (0; 200; 400 e 600 mg L⁻¹), distribuídos em delineamento de blocos casualizados, em esquema parcelas subdivididas com três repetições, cada parcela foi constituída por três plantas úteis. O déficit hídrico na fase vegetativa/frutificação afetou de forma negativa o crescimento em diâmetro de caule, volume de copa, diâmetro de copa e índice de vigor vegetativo, peso médio de frutos, diâmetro polar e equatorial dos frutos da goiabeira, aos 390 dias após o transplantio. A aplicação foliar de ácido ascórbico na concentração de 600 mg L⁻¹ proporcionou efeitos positivos no peso total de frutos e produtividade da goiabeira, aos 390 dias após o transplantio. O déficit hídrico na fase de maturação estimulou a síntese de vitamina C e sólidos solúveis da goiabeira aos 390 dias após o transplantio.

Palavras-chave: *Psidium guajava* L., composto não-enzimático, déficit hídrico.

INTRODUCTION

Guava (*Psidium guajava* L.) is a species native to tropical America, widely distributed throughout the Brazilian territory, which stands out for producing fruits with a pleasant flavor, high nutritional value and great acceptance in the market

(SILVA et al., 2015). The fruits can be consumed fresh or processed into various products, such as guava paste, jellies, pulps, fruits in syrup, puree, bases for drinks, soft drinks, juices and syrups (COSTA et al., 2017). The versatility and nutritional value of the fruits make guava a fruit crop of great socioeconomic importance in Brazil (OLIVEIRA et al., 2019).

In the 2023 harvest, Brazil reached an average production of 582,832 tons spread over 22,487 hectares, resulting in an average yield of 25,919 kg per hectare, with the Northeast region standing out as one of the main producers, with the states of Pernambuco, Bahia and Ceará leading with productions of 205,960, 45,135 and 39,493 tons, respectively; Paraíba contributed approximately 2,692 tons, which is equivalent to an average yield of 7,237 kg per hectare (IBGE, 2024).

Although, this fruit crop has great potential in the region, its production is limited by water restriction. This is due to the inconstancy of precipitation, in terms of both quantity and spatial distribution, with most of the rain concentrated in just four months of the year in several municipalities (NÓBREGA et al., 2024). The decrease in water availability can cause negative effects on plant growth, biochemical properties, yield, and quality of fruits (FÁTIMA et al., 2024a). However, the intensity of this damage varies according to the species, cultivar, edaphoclimatic conditions, management practices, time of exposure to stress, in addition to the phenological stage of the plant (ARAÚJO et al., 2010).

Water deficit irrigation management consists of applying the amount of water necessary to maintain agricultural production, allowing plants to go through short periods of water stress (OLIVEIRA et al., 2020). According to SILVA et al. (2018), this method can reduce water consumption, improve fruit quality, and increase plant resistance to drought conditions. To maximize the potential of guava cultivation in the semi-arid region of Brazil, it is essential to establish irrigation strategies adapted to local climatic conditions to maximize its production.

Thus, foliar application of elicitors has been widely studied, due to their role as abiotic stress-signaling agents, contributing to the acclimatization of plants under water restriction (WANG et al., 2018). Among the elicitors, ascorbic acid (AsA) stands out for its function as an antioxidant agent, protecting plants against oxidative stress. AsA acts in the detoxification of reactive oxygen species (ROS) and in the protection of lipids and proteins against the adverse effects of oxidative stress, caused by water deficit (AKRAM et al., 2017; SHARMA et al., 2019).

The mitigating action of ascorbic acid has been observed in several studies, highlighting its role in the regulation of antioxidant defense processes in plants under water deficit conditions, as demonstrated by FÁTIMA et al. (2024b) in sour passion fruit (*Passiflora edulis* Sims) and by LACERDA et al.

(2025) and TORRES et al. (2025) in guava (*Psidium guajava* L.).

In view of the above, this study evaluated the effects of foliar application of ascorbic acid on the morphophysiology, production components and postharvest quality of guava fruits, cv. Paluma, under irrigation strategies with water deficit in a semi-arid area.

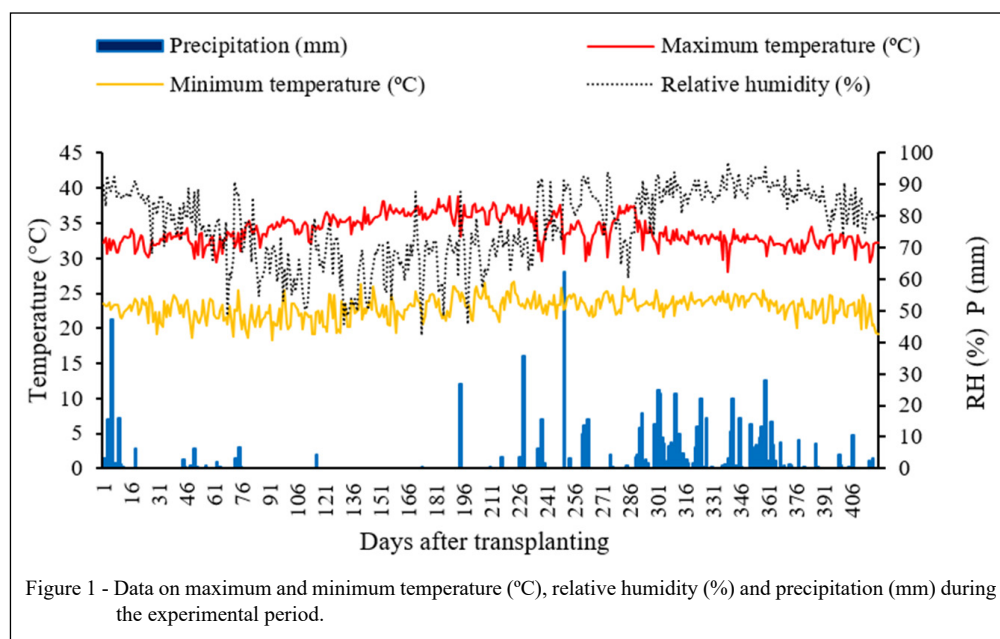
MATERIALS AND METHODS

The experiment was carried out under field conditions in the area belonging to the 'Rolando Enrique Rivas Castellón' Experimental Farm of the Center for Sciences and Agri-Food Technology - CCTA of the Universidade Federal de Campina Grande - UFCG, located in the municipality of São Domingos, Paraíba, Brazil (06°48'50"S; 37°56'31"W, 190 m). According to Köppen's classification, the climate of the region is BSh, which means a hot semi-arid climate, with scarce rainfall and irregular distribution throughout the year (ALVARES et al., 2013). The climate data collected during the experiment, temperature (maximum and minimum), relative humidity, and precipitation, are shown in figure 1.

Treatments consisted of the combination of seven irrigation strategies with water deficit - IMS (SE - plants under full irrigation throughout the cycle - 100% crop evapotranspiration - ET_c; EV - plants irrigated with 50% ET_c in the vegetative stage; FL - plants under water deficit in the flowering stage; FR - fruiting stage; VE/FL - in the vegetative and flowering stages, with full irrigation in the fruiting stage; VE/FR - plants subjected to water deficit in the vegetative and fruiting stages) and four concentrations of ascorbic acid - AsA (0, 200, 400 and 600 mg L⁻¹), distributed in a randomized block design, in a split-plot scheme with three replicates, each plot consisting of 3 usable plants and a 3 m border between the plots. Plots corresponded to irrigation strategies and subplots to ascorbic acid concentrations. Ascorbic acid concentrations were defined based on a study conducted by GAAFAR et al. (2020) using 0, 200 and 400 mg L⁻¹ in common bean under water deficit conditions.

The cultivar used in this study was Paluma, propagated by cuttings. Paluma is a cultivar that has large fruits (over 200 g), with a piriform shape, a short 'neck' and smooth skin. Its pulp is thick (from 1.3 cm to 2.0 cm), firm, with intense red color and pleasant flavor (EMBRAPA, 2010).

The area was prepared by plowing followed by harrowing, aiming the unraveling and



leveling of the area. The soil of the experimental area is classified as *Neossolo Flúvico Ta Eutrófico típico* (Fluvent) of loamy sand texture. Before transplanting the seedlings to the field, soil samples were collected in the experimental area, in the 0-40 cm layer, and then mixed to form a composite sample, whose chemical and physical characteristics were obtained according to the methodology of TEIXEIRA et al. (2017) and are presented in table 1.

The guava seedlings were transplanted to the field in 30 cm × 30 cm × 30 cm holes when they reached 40 cm in height. At transplanting, the collar (region between the roots and the trunk) of the seedlings was above the ground level, to avoid the

emergence of roots. Formative pruning was carried out to guide the plant to obtain well-distributed branches with balanced and airy architecture, thus allowing a greater penetration of light and favoring ventilation inside the crown. The plants were conducted on a single stem up to 50 cm in height, when the terminal bud was eliminated. From the shoots that appeared, four well-distributed branches were left in the directions of the four cardinal points. After reaching maturity, the main branches were pruned, so that they remained with a length of 40 cm.

Fertilization with nitrogen, phosphorus and potassium was carried out as recommended by BARBOSA & LIMA (2010). Fertilization with P_2O_5

Table 1 - Physical-hydraulic and chemical attributes of the soil (0–0.30m) used in the experiment.

Chemical characteristics ^a								
pH (H ₂ O)	OM	P	K ⁺	Na ⁺	Ca ²⁺	Mg ²⁺	Al ³⁺	H ⁺
(1:2.5)	g kg ⁻¹	(mg kg ⁻¹)	cmol _c kg ⁻¹					
7.19	1.40	5.95	0.49	0.07	4.70	3.63	0	0
Chemical characteristics				Physical characteristics				
EC _{se}	CEC	SAR	ESP	Particle-size fraction (g kg ⁻¹)			Moisture (dag kg ⁻¹)	
(dS m ⁻¹)	cmol _c kg ⁻¹	(mmol L ⁻¹) ^{0.5}	%	Sand	Silt	Clay	33.42 kPa ¹	1519.5 kPa ²
0.58	8.89	1.40	0.79	73.51	20.14	6.35	15.78	6.41

Determined attributes: pH - Hydrogen potential, OM – Organic matter: Walkley-Black Wet Digestion; Ca²⁺ and Mg²⁺ extracted with 1 M KCl at pH 7.0; Na⁺ and K⁺ extracted with 1 M NH₄OAc at pH 7.0; Al³⁺+H⁺ extracted with 0.5 M CaOAc at pH 7.0; EC_{se} - Electrical conductivity of the saturation extract; Estimated attributes: CEC - Cation exchange capacity; SAR_{se} - Sodium adsorption ratio of the saturation extract; ESP - Exchangeable sodium percentage.

and K_2O was calculated considering the potential fertility of the soil. Fertilization with micronutrients was carried out every 15 days to avoid the appearance of nutritional deficiencies. In the vegetative stage, nitrogen fertilization was split into portions and supplied in 24 applications per year, starting at 30 days after transplanting. In the flowering and fruiting stages, 30% of the nitrogen was supplied after fruiting pruning, and the remaining 70% was applied in a decreasing manner, up to 150 days after pruning, as described by BARBOSA & LIMA (2010).

Potassium fertilization was applied 30% after pruning, 15% after fruit set, 25% in the intermediate stage of fruit growth and 30% in the final stage of fruit growth (before maturation). Phosphate fertilization was carried out at once at planting and before each fruiting pruning, considering the results of the soil analysis. Cultural practices and phytosanitary treatments were carried out as recommended for the crop, by monitoring the emergence of pests and diseases and adopting control measures when necessary.

The irrigation system adopted was localized by drip, using 32 mm PVC pipes in the main line and 16 mm low-density polyethylene pipes in the lateral lines, with drippers with flow rate of 10 L h⁻¹. Two pressure-compensating drippers (GA 10 Gripa model) were installed for each plant, each one 15 cm away from the stem. The plants were irrigated daily, at 07:00 am, with water supply according to the strategy adopted, and the depth was estimated based on ET_c according to BERNARDO et al. (2019), using Eq. 1:

$$ET_c = ET_o \times K_c \quad (1)$$

Where:

ET_c - Crop evapotranspiration, mm d⁻¹;

ET_o - Reference evapotranspiration, mm d⁻¹;

K_c = crop coefficient,

The K_c values adopted were 0.40 until flowering and 0.65 from flowering to the end of the cultivation cycle, according to EMBRAPA (2010).

Soil moisture content was also quantified, using the Time Domain Reflectometer (TDR), through high-frequency electromagnetic pulse emissions by the sensor connected to a coaxial cable, which reads the reflections occurred during a given time and read them as distance, according to their relationship with the dielectric constant (TOPP et al., 1980). In the experimental area, access pipes were installed in each treatment, totaling 16 treatments, distributed throughout the experiment, so the TDR was previously calibrated in the cultivation area, and readings were taken weekly before the beginning of irrigation.

Macronutrients were applied by fertigation using a Venturi injector at 15-day intervals after transplanting (CAVALCANTI, 2008), and their quantity was established based on soil analysis. Fertilization with micronutrients was performed every 30 days starting at 30 DAT, by foliar application, with a solution at a concentration of 1.0 g L⁻¹ of Dripsol Micro® (1.2% magnesium, 0.85% boron, 3.4% iron, 4.2% zinc, 3.2% manganese, 0.5% copper and 0.06% molybdenum).

Foliar applications with ascorbic acid began at 72 h before the imposition of water deficit on the plants. Subsequently, the applications were carried out every 20 days and started at 5 pm until the plants reached the full flowering stage. Ascorbic acid concentrations were obtained by dissolving it in distilled water, which was prepared on the day of each application. AsA was applied by foliar applications, to fully wet the aerial part. A surfactant (Tween 20 to 0.025%) was added to the spray solutions to break the surface tension of the water and favor the penetration of ascorbic acid into the leaves. The plants were isolated with a structure made of plastic tarpaulin during the applications of ascorbic acid concentrations to prevent the solutions from drifting. The average volume of spray applied during the experimental period was 1.256 L per plant.

At 390 days after transplanting (DAT), stem diameter was determined using a digital caliper; crown diameter (DCrown) was obtained through the average of the crown diameter observed in the row direction (RD) and interrow direction (IRD); crown volume (VCrown) was calculated from plant height (H), RD and IRD, using Eq. 2, and VVI was calculated according to Portella et al. (2016), using Eq. 3:

$$V_{Crown} = \left(\frac{\pi}{6}\right) \times H \times RS \times IRD \quad (2)$$

Where:

VCrown - crown volume (m³);

H - plant height (m);

RD - crown diameter in the row direction (m); and

IRD - diameter of the crown in the interrow direction (m).

$$VVI = \frac{[H + D_{Crown} + (SD \times 10)]}{100} \quad (3)$$

Where:

VVI - vegetative vigor index;

H - plant height (m);

DCrown - crown diameter (m); and

SD - stem diameter (m).

As production components, the following parameters were measured: number of fruits per plant (NF), average fruit weight (AFW), total fruit weight (TFW), and yield (YLD). The fruits were

harvested when they showed a change from green to yellow, considered to be the point of harvest. NF was determined by counting the fruits harvested. TFW was obtained by the sum of the weight of all fruits produced per plant. The AFW was obtained through the ratio between the fresh mass of fruits and the total number of fruits harvested. Fruit diameters were obtained with a digital caliper, and yield was obtained considering the planting density of the area and extrapolated to one hectare.

For postharvest quality, the following parameters were evaluated: physical attributes, polar (PD) and equatorial (ED) diameters, and the chemical characteristics of the fruits, through the hydrogen potential - pH, soluble solids - SS ($^{\circ}$ Brix), titratable acidity - TA (% citric acid) and ascorbic acid - AA ($\text{mg } 100\text{g}^{-1}$ of pulp).

Guava pulp pH was determined using a digital potentiometer with a glass membrane electrode and a resolution of 0.01. Soluble solids were expressed in $^{\circ}$ Brix and determined using a portable refractometer with a resolution of 0.2. Titratable acidity was measured according to the standards of the Adolfo Lutz Institute (IAL, 2008) and expressed as a percentage of citric acid. The concentration of ascorbic acid was determined by titration, using 1 g of pulp, plus 49 mL of 0.5% oxalic acid and titrated against Tillmans' solution until the appearance of pink color (IAL, 2008).

The data collected in this study were subjected to the normality test (Shapiro-Wilk) and subsequently subjected to analysis of variance using the F test and, when there was significance, the mean comparison test (Scott-Knott) was performed at the 0.05 probability level for the factor irrigation strategies with water deficit, and linear and quadratic polynomial regression analysis for the different concentrations of ascorbic acid, using the statistical software SISVAR-ESAL version 5.7 (FERREIRA, 2019).

The multivariate structure of the results was evaluated by principal component analysis (PCA), synthesizing the amount of relevant information contained in the original data set in a smaller number of dimensions, resulting from linear combinations of the original variables generated from the eigenvalues ($\lambda \geq 1.0$) in the correlation matrix, explaining a percentage greater than 10% of the total variance (GOVAERTS et al., 2007).

From the reduction of the dimensions, the original data of the variables of each component were subjected to multivariate analysis of variance (MANOVA) by the HOTELLING's (1947) test at 0.05 probability level for irrigation strategies with

water deficit and ascorbic acid concentrations, as well as for the interaction between the factors. Variables with a correlation coefficient greater than or equal to 0.65 were maintained in each principal component (PC) (HAIR et al., 2009). Statistical analyses were performed using Statistica v.7.0 software (STATSOFT, 2024).

RESULTS AND DISCUSSION

There were significant effects for the interaction between the factors irrigation management strategies and foliar application of ascorbic acid (IMS $\times$ AsA) on the polar fruit diameter (PD), equatorial fruit diameter (ED), number of fruits (NF), average fruit weight (AFW), total fruit weight (TFW), and yield (YLD) of guava (Table 2). Irrigation management strategies had a significant effect on the relative water content (RWC) of guava 390 days after transplanting.

There was interaction between the factors irrigation management strategies and foliar application of ascorbic acid (IMS $\times$ AsA), showing significant effects on canopy volume (VCrown), canopy diameter (DCrown), vegetative vigor index (VVI), vitamin C (Vit-C), soluble solids (SS), hydrogen potential (pH), and titratable acidity (TA) of guava fruits (Table 3). There was a significant effect of ascorbic acid concentrations on stem diameter (SD) of guava trees at 390 days after transplanting.

The multidimensional space of the original variables was reduced to two principal components (PC1 and PC2) with eigenvalues greater than $\lambda \geq 1.0$, as highlighted by Kaiser (1960). The eigenvalues and percentage of variance explained for each component (Table 4) together represent 50.69% of the total variation. PC1 explained 27.74% of the total variance, and PC2 represented 22.95% of the variance.

A significant effect ($P \leq 0.01$) of the interaction between irrigation management strategies (IMS) with water deficit and ascorbic acid (AsA) concentrations was observed for PC1 and PC2 (Table 4). A significant effect ($P \leq 0.01$) of irrigation management strategies and ascorbic acid concentrations was also found when analyzed separately.

The multidimensional space of the original variables was reduced to two principal components (PC1 and PC2) with eigenvalues greater than $\lambda \geq 1.0$, as highlighted by Kaiser (1960). The eigenvalues and percentage of variance explained for each component (Table 5) together represent 50.69% of the total variation. PC1 explained 27.74% of the total variance, and PC2 represented 22.95% of the variance.

Table 2 - Summary of analysis of variance for the polar fruit diameter (PD), equatorial fruit diameter (ED), number of fruits (NF), average fruit weight (AFW), total fruit weight (TFW), yield (YLD), relative water content (RWC), and electrolyte leakage (EL%) of guava grown under irrigation management strategies and foliar application of ascorbic acid, at 390 days after transplanting.

Sources of variation	DF	-----Mean squares-----							
		TFW	AFW	NF	PD	ED	YLD	RWC	EL%
IMS	5	3.11 ^{ns}	2443.46*	150.12*	220.61*	65.49*	4055098.58*	112.27*	16.78 ^{ns}
Residual 1	10	1.94	22.15	5.41	3.60	2.02	446930.72	11.16	12.56
AsA	3	2.80*	1710.31*	105.67*	9.28 ^{ns}	12.59 ^{ns}	3672915.72*	75.78 ^{ns}	7.54 ^{ns}
Linear. R	1	0.91 ^{ns}	2080.22*	1.53 ^{ns}	0.27 ^{ns}	37.49*	1183346.24 ^{ns}	41.71 ^{ns}	0.27 ^{ns}
Quadratic. R	1	1.56*	52.94 ^{ns}	43.38*	14.02*	0.00 ^{ns}	2051115.82*	15.83 ^{ns}	2.95 ^{ns}
IMS × AsA	15	6.70*	723.38*	209.97*	92.67*	25.39*	8764269.60*	29.67 ^{ns}	17.77 ^{ns}
Residual 2	36	0.35	19.88	3.27	8.75	7.17	462483.76	28.71	15.87
Blocks	2	1.94 ^{ns}	34.76 ^{ns}	33.34 ^{ns}	27.05 ^{ns}	2.47 ^{ns}	2534477.47 ^{ns}	11.16 ^{ns}	13.64 ^{ns}
CV 1 (%)		8.90	2.70	6.53	2.34	2.21	8.90	6.48	20.46
CV 2 (%)		9.07	2.56	5.08	3.64	4.16	9.06	7.48	23.00

IMS - irrigation management strategy, AsA - ascorbic acid concentrations.

* and **significant at the probability of 0.05 and 0.01 by the F test, respectively; ns not significant by the F test. CV = coefficient of variation; GL - Degree of freedom.

A significant effect ($P \leq 0.01$) of the interaction between irrigation management strategies (IMS) with water deficit and ascorbic acid (AsA) concentrations was observed for PC1 and PC2 (Table 5). A significant effect ($P \leq 0.01$) of irrigation management strategies and ascorbic acid concentrations was also found when analyzed separately.

The behavior of the treatments is expressed in figure 2A and figure 2B, referring to the first and second principal components (PC1 and PC2). In the

first principal component (PC1) there is possibly an interaction between irrigation strategies and ascorbic acid concentrations. It can be observed that the correlation coefficients were greater than 0.60 for the variables TFW, PD, RWC, EL%, SD, VCrown, DCrown, VVI, YLD, and SS.

In the principal component 1, for stem diameter (SD), crown volume (VCrown), crown diameter (DCrown) and vegetative vigor index (VVI), the highest values (44.10 mm, 6.87 m³, 2.86 m² and

Table 3 - Summary of the analysis of variance regarding stem diameter (SD), canopy volume (VCrown), canopy diameter (DCrown), vegetative vigor index (VVI), vitamin C (Vit-C), soluble solids (SS), hydrogen potential (pH), titratable acidity (TA), of guava trees cultivated under irrigation management strategies and foliar application of ascorbic acid, 390 days after transplanting.

Sources of variation	DF	-----Mean squares-----							
		SD	VCrown	DCrown	VVI	Vit-C	SS	pH	TA
IMS	5	41.12 ^{ns}	5.85*	0.12*	0.00*	35.62*	35.62*	0.009*	0.23*
Residual 1	10	16.14	0.71	0.02	0.00	3.77	3.77	0.000	0.03
AsA	3	33.25*	6.91*	0.13*	0.00*	18.97*	18.97*	0.003*	0.01 ^{ns}
Linear. R	1	14.83 ^{ns}	8.16*	0.16*	0.00*	5.62 ^{ns}	5.62 ^{ns}	0.001*	0.01 ^{ns}
Quadratic. R	1	83.85*	9.22*	0.21*	0.00*	45.95*	45.95*	0.002*	0.00*
IMS × AsA	15	18.77 ^{ns}	1.36*	0.04*	0.00*	23.65*	23.65*	0.005*	0.11*
Residual 2	36	11.96	0.60	0.01	19.88	2.66	2.66	0.000	0.03
Blocks	2	37.51	1.14 ^{ns}	0.01 ^{ns}	0.00 ^{ns}	12.47 ^{ns}	12.47 ^{ns}	0.000 ^{ns}	0.05 ^{ns}
CV 1 (%)		9.99	17.34	5.81	8.57	5.37	5.37	0.26	7.52
CV 2 (%)		8.60	15.98	5.20	6.52	4.51	4.51	0.23	7.03

IMS - irrigation management strategy, AsA - ascorbic acid concentrations.

* and ** significant at the probability of 0.05 and 0.01 by the F test, respectively; ns not significant by the F test.

CV = coefficient of variation; GL - Degree of freedom.

Table 4 - Eigenvalues, percentage of total variance explained, in the multivariate analysis of variance (MANOVA) and the correlation coefficients (r) between the original variables and the principal components.

								Principal components (PCs)	
								PC1	PC2
Eigenvalues (λ)								4.63	3.67
Percentage of total variance (S2%)								27.74	22.95
Hotelling test (T2) for irrigation management strategy (IMS)								0.01	0.01
Hotelling test (T2) for ascorbic acid (AsA)								0.01	0.01
Hotelling test (T2) for interaction (IMS $\times$ AsA)								0.01	0.01
-----Correlation coefficient-----									
PCs	TFW	AFW	NF	PD	ED	YLD	RWC	EL%	
PC1	0.67*	0.16	0.60	0.60*	0.45	0.30	0.62*	0.62*	
PC2	0.48	0.44*	0.64*	0.26	0.62*	0.64*	0.39	0.17	
-----Mean values-----									
	TFW	AFW	NF	PD	ED	YLD	RWC	EL%	
T1 - SEA1	9.594	203.499	49.259	77.473	61.340	10965.079	76.848	14.216	
T2 - SEA2	5.990	205.168	31.037	86.856	66.783	6845.714	75.163	17.298	
T3 - SEA3	6.282	159.577	35.444	77.910	66.588	7179.683	70.463	19.165	
T4 - SEA4	6.453	165.427	31.000	79.184	61.971	7375.238	72.320	13.112	
T5 - FRA1	6.971	144.665	44.222	84.454	61.339	7966.984	73.177	15.795	
T6 - FRA2	5.096	163.575	25.000	79.793	62.938	5823.492	77.924	18.195	
T7 - FRA3	7.807	164.813	36.852	77.782	60.714	8921.905	71.794	15.840	
T8 - FRA4	4.296	162.331	33.037	75.566	63.032	4909.206	74.651	15.113	
T9 - VE/FRA1	6.640	153.189	42.037	75.986	61.662	7588.571	72.633	18.706	
T10 - VE/FRA2	7.538	172.402	41.889	79.512	65.990	8614.603	75.832	17.157	
T11 - VE/FRA3	8.554	133.871	46.148	71.567	58.383	9776.508	77.437	15.151	
T12 - VE/FRA4	6.098	153.192	34.148	75.088	61.787	6968.889	71.319	21.142	
T13 - VEA1	5.913	191.875	28.963	78.121	67.065	6758.095	69.926	16.438	
T14 - VEA2	8.017	203.614	39.704	84.046	66.101	9161.905	76.551	16.510	
T15 - VEA3	7.666	171.309	48.593	77.255	64.707	8760.635	71.480	19.210	
T16 - VEA4	5.211	155.024	32.259	80.201	61.700	5955.556	69.141	15.065	
T17 - VE/FLA1	6.007	171.676	30.333	81.326	68.440	6864.762	70.821	18.144	
T18 - VE/FLA2	4.468	202.478	23.037	86.245	63.431	5106.032	70.841	19.833	
T19 - VE/FLA3	6.353	178.837	35.593	98.943	73.357	7260.952	59.092	18.628	
T20 - VE/FLA4	8.552	193.202	42.148	83.702	67.220	9773.968	71.290	19.341	
T21 - FLA1	5.080	200.492	20.741	86.010	67.870	5805.714	65.769	15.608	
T22 - FLA2	7.180	168.863	42.556	77.198	62.992	8205.714	70.890	18.718	
T23 - FLA3	5.656	174.964	31.111	83.546	66.620	6463.492	68.125	18.159	
T24 - FLA4	6.246	184.314	30.185	92.954	65.056	7137.778	66.434	19.260	
T25 - MATA1	4.302	163.965	22.889	75.940	65.933	4916.825	61.642	19.196	
T26 - MATA2	4.924	181.557	20.111	81.468	66.008	4985.079	67.033	16.338	
T27 - MATA3	5.041	183.164	26.185	78.763	65.749	5761.270	67.176	17.158	
T28 - MATA4	4.798	199.708	22.148	83.422	65.978	5483.175	68.130	17.451	

T - Treatment; SE - plants under full irrigation throughout the cycle; VE - plants irrigated with water deficit (50% crop evapotranspiration - ETC) in the vegetative stage; FL - in the flowering stage; FR - in the fruiting stage; MAT - in the fruit maturation stage; VE/FL - in the vegetative and flowering stages; VE/FR - in the vegetative and fruiting stages); A - Ascorbic acid - A1 (0 mg L⁻¹); A2 (200 mg L⁻¹); A3 (400 mg L⁻¹); A4 (600 mg L⁻¹); TFW (Total fruit weight/plant - kg); AFW (Average fruit weight - g per fruit); PD (Fruit polar diameter - mm); ED (Fruit equatorial diameter - mm); YLD (Yield - kg ha⁻¹); RWC (Relative water content - %); EL% (Electrolyte leakage - %).

0.09, respectively) were observed in plants subjected to water deficit in the fruiting stage and without application of ascorbic acid (T5), when compared to plants grown under 50% ETC in the vegetative/fruiting stages (T9).

Among the strategies, the intermittent water deficit, that is, in the vegetative stage and later in the fruiting stage, contributed to inhibiting plant growth, which was already explained by the deleterious

Table 5 - Eigenvalues, percentage of total variance explained, in the multivariate analysis of variance (MANOVA) and the correlation coefficients (r) between the original variables and the principal components.

						Principal components (PCs)		
						PC1	PC2	
Eigenvalues (λ)						4.63	3.67	
Percentage of total variance (S2%)						27.74	22.95	
Hotelling test (T2) for irrigation management strategy (IMS)						0.01	0.01	
Hotelling test (T2) for ascorbic acid (AsA)						0.01	0.01	
Hotelling test (T2) for interaction (IMS $\times$ AsA)						0.01	0.01	
-----Correlation coefficient-----								
PCs	SD	VCrown	DCrown	VVI	Vit-C	SS	pH	TA
PC1	0.70*	0.70*	0.63*	0.71*	0.09	0.52*	0.19	0.01
PC2	0.58	0.62	0.62	0.65	0.30*	0.18	0.61*	0.60*
-----Mean values-----								
	SD	VCrown	DCrown	VVI	Vit-C	SS	pH	TA
T1 - SEA1	43.977	6.414	2.767	0.088	43.387	12.600	4.273	2.134
T2 - SEA2	40.190	4.733	2.475	0.080	37.066	10.633	4.417	2.494
T3 - SEA3	43.640	6.134	2.683	0.087	35.917	11.200	4.367	2.578
T4 - SEA4	43.737	5.885	2.675	0.086	40.514	10.700	4.437	2.747
T5 - FRA1	44.100	6.867	2.858	0.090	38.215	11.367	4.427	2.768
T6 - FRA2	39.120	4.888	2.683	0.079	35.055	12.900	4.387	2.663
T7 - FRA3	43.353	5.496	2.550	0.085	35.917	11.833	4.450	2.494
T8 - FRA4	41.633	5.532	2.692	0.083	31.607	11.733	4.387	2.684
T9 - VE/FRA1	35.660	3.751	2.342	0.072	36.779	13.033	4.417	2.980
T10 - VE/FRA2	40.540	3.841	2.342	0.077	37.066	10.567	4.423	2.494
T11 - VE/FRA3	37.247	4.170	2.450	0.075	36.779	12.067	4.370	2.874
T12 - VE/FRA4	41.107	4.780	2.542	0.081	35.917	12.500	4.380	2.726
T13 - VEA1	40.307	6.286	2.750	0.084	38.215	11.967	4.380	2.663
T14 - VEA2	39.530	4.575	2.517	0.078	35.055	10.267	4.443	2.557
T15 - VEA3	36.017	3.841	2.375	0.073	37.353	12.133	4.393	2.705
T16 - VEA4	36.960	3.633	2.325	0.073	38.503	12.233	4.430	2.853
T17 - VE/FLA1	43.313	5.088	2.658	0.084	38.332	10.900	4.403	2.515
T18 - VE/FLA2	35.763	3.805	2.383	0.078	37.221	11.933	4.417	2.282
T19 - VE/FLA3	36.940	4.165	2.575	0.075	35.276	11.900	4.457	2.451
T20 - VE/FLA4	41.110	4.668	2.558	0.080	31.943	10.900	4.383	2.409
T21 - FLA1	45.407	6.215	2.725	0.089	37.928	12.600	4.303	2.304
T22 - FLA2	39.923	4.373	2.542	0.078	35.055	12.067	4.383	2.599
T23 - FLA3	37.400	4.071	2.450	0.075	29.883	11.967	4.327	2.536
T24 - FLA4	39.253	3.553	2.425	0.075	40.083	13.067	4.357	2.451
T25 - MATA1	38.390	4.396	2.533	0.077	47.697	14.100	4.330	2.409
T26 - MATA2	40.023	5.093	2.525	0.081	37.928	13.667	4.313	2.811
T27 - MATA3	40.683	4.897	2.550	0.081	37.641	13.367	4.323	2.790
T28 - MATA4	35.967	3.996	2.323	0.073	40.514	12.500	4.287	2.473

T - Treatment; SE - plants under full irrigation throughout the cycle; VE - plants irrigated with water deficit (50% crop evapotranspiration - ETc) in the vegetative stage; FL - in the flowering stage; FR - in the fruiting stage; MAT - in the fruit maturation stage; VE/FL - in the vegetative and flowering stages; VE/FR - in the vegetative and fruiting stages); A - Ascorbic acid - A1 (0 mg L⁻¹); A2 (200 mg L⁻¹); A3 (400 mg L⁻¹); A4 (600 mg L⁻¹); SD (Stem diameter - mm); VCrown (Crown volume - m³); DCrown (Crown diameter - m²); VVI (Vegetative Vigor Index); Vit-C (Vitamin C - mg 100 g⁻¹ of pulp); SS (Soluble Solids - °Brix); pH (hydrogen potential) and TA (titratable acidity -% citric acid).

effects of water deficit that vary according to the intensity, duration and development stage in which the plants were subjected (ARAÚJO et al., 2010). In

a study carried out with sour passion fruit, FÁTIMA et al. (2024b) found greater tolerance to water deficit in the flowering stage, and according to the authors,

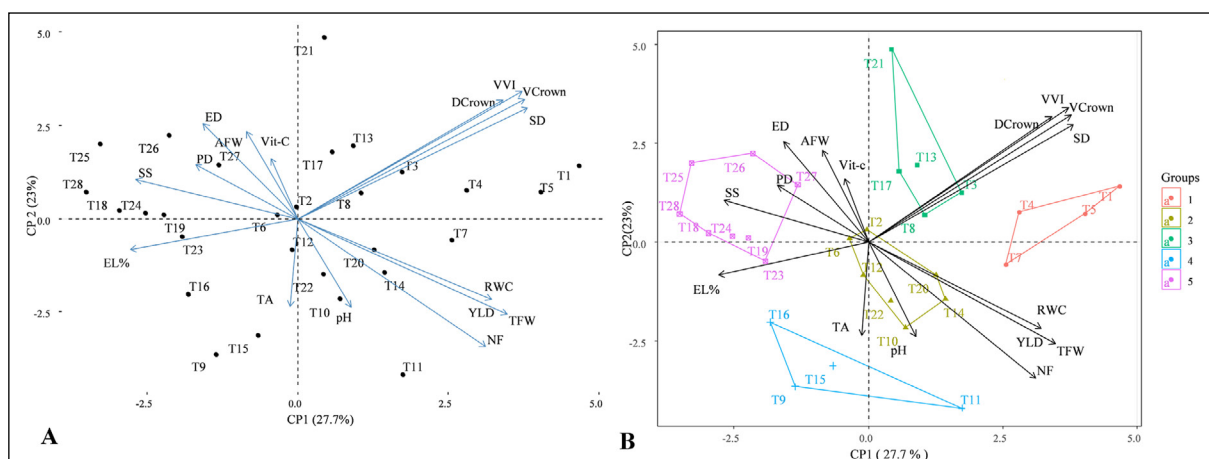


Figure 2 - Two-dimensional projection of the scores of the principal components for the factors irrigation strategies with water deficit - IMS and concentrations of ascorbic acid - AsA (A) and of the variables analyzed (B) in the two principal components (PC1 and PC2).

T1- plants under full irrigation throughout the cycle and without application of ascorbic acid; T2- plants under full irrigation throughout the cycle and application of 200 mg L⁻¹ of ascorbic acid; T3- plants under full irrigation throughout the cycle and application of 400 mg L⁻¹ of ascorbic acid; T4- plants under full irrigation throughout the cycle and application of 600 mg L⁻¹ of ascorbic acid; T5 - water deficit (50% of crop evapotranspiration - ETc) in the vegetative stage and without application of ascorbic acid; T6 - water deficit (50% of crop evapotranspiration - ETc) in the vegetative stage and application of 200 mg L⁻¹ of ascorbic acid; T7 - water deficit (50% of crop evapotranspiration - ETc) in the vegetative stage and application of 400 mg L⁻¹ of ascorbic acid; T8 - water deficit (50% of crop evapotranspiration - ETc) in the vegetative stage and application of 600 mg L⁻¹ of ascorbic acid; T9 - in the flowering stage and without application of ascorbic acid; T10 - in the flowering stage and application of 200 mg L⁻¹ of ascorbic acid; T11 - in the flowering stage and application of 400 mg L⁻¹ of ascorbic acid; T12 - in the flowering stage and application of 600 mg L⁻¹ of ascorbic acid; T13 - in the fruiting stage and without application of ascorbic acid; T14 - in the fruiting stage and application of 200 mg L⁻¹ of ascorbic acid; T15 - in the fruiting stage and application of 400 mg L⁻¹ of ascorbic acid; T16 - in the fruiting stage and application of 600 mg L⁻¹ of ascorbic acid; T17 - in the maturation stage and without application of ascorbic acid; T18 - in the maturation stage and application of 200 mg L⁻¹ of ascorbic acid; T19 - in the maturation stage and application of 400 mg L⁻¹ of ascorbic acid; T20 - in the maturation stage and application of 600 mg L⁻¹ of ascorbic acid; T21 - in the vegetative and flowering stages and without application of ascorbic acid; T22 - in the vegetative and flowering stages and application of 200 mg L⁻¹ of ascorbic acid; T23 - in the vegetative and flowering stages and application of 400 mg L⁻¹ of ascorbic acid; T24 - in the vegetative and flowering stages and application of 600 mg L⁻¹ of ascorbic acid; T25 - in the vegetative and fruiting stages and without application of ascorbic acid; T26 - in the vegetative and fruiting stages and application of 200 mg L⁻¹ of ascorbic acid; T27 - in the vegetative and fruiting stages and application of 400 mg L⁻¹ of ascorbic acid; T28 - in the vegetative and fruiting stages and application of 600 mg L⁻¹ of ascorbic acid; TFW (Total fruit weight/plant - kg); AFW (Average fruit weight - g per fruit); PD (Fruit polar diameter - mm); ED (Fruit equatorial diameter - mm); YLD (Yield - kg ha⁻¹); RWC (Relative water content - %); EL% (Electrolyte leakage - %); SD (Stem diameter - mm); VCrown (Crown volume - m³); DCrown (Crown diameter - m²); VVI (Vegetative Vigor Index); Vit-C (Vitamin C - mg 100 g⁻¹ of pulp); SS (Soluble Solids - °Brix); pH (Hydrogen potential) and TA (Titratable Acidity -% Citric Acid).

the water deficit caused changes in photosynthetic activities at different stages of development of sour passion fruit plants.

Plant growth inhibition under water deficit conditions may be associated with the reduction of turgor pressure in cells. Turgor pressure is critical for cell rigidity and thus for cell expansion and growth; when water availability is low, turgor pressure decreases, resulting in limitations for cell division and expansion (ANJUM et al., 2016). Under conditions of water deficit, growth inhibition results from the diversion of metabolic energy to maintain the vital functions for plant development, such as the synthesis of organic and inorganic solutes (FÁTIMA et al., 2019).

The highest value (77.92%) for relative water content in the leaf blade (RWC) was observed in plants grown under water deficit (50% ETc) in the fruiting stage and ascorbic acid concentration of 200 mg L⁻¹ (T6), while the lowest value was obtained under water deficit in the vegetative/flowering stages and ascorbic acid concentration of 400 mg L⁻¹ (T19). Regarding electrolyte leakage (EL%), the lowest value (13.11%) was found in plants irrigated with 100% ETc during the entire cycle and under ascorbic acid concentration of 600 mg L⁻¹ (T4).

The increase in the relative water content with the application of ascorbic acid may be related to its performance in improving turgor potential, which promotes the maintenance of water balance in plant

tissues (EL-BIALLY et al., 2018). AsA also promotes increase in cell wall strength and in its ability to withstand water loss under abiotic stress conditions such as water stress (NAZ et al., 2016).

The reduction in electrolyte leakage observed when the ascorbic acid concentration of 600 mg L⁻¹ was applied to guava cultivated under water deficit in the fruiting stage may be indicative of the elimination of ROS and protection of lipids and proteins (FATAH & SADEK, 2020). Usually under conditions of water deficit, there tends to be an increase in the production of ROS, such as hydroxyl radicals and lipid peroxides, which can contribute to the degradation of membrane lipids, causing the breakdown of lipid structures and impairing the function of membranes (PEIFANG et al., 2015).

Regarding the total fruit weight (TFW) and yield (YLD) of guava plants, it was found that full irrigation (100% ETc) throughout the cycle and without application of ascorbic acid (T1), promoted higher TFW of 9.59 kg per plant and YLD of 10.97 ton ha⁻¹ when compared to the T26 treatment, i.e., plants under water deficit in the fruiting stage and application of ascorbic acid at a concentration of 600 mg L⁻¹ showed increments of 50.20 and 55.22%, respectively. The yield obtained in this study is above the average produced by the state of Paraíba, which is 7.24 ton ha⁻¹, according to IBGE (2024).

The reduction in total fruit weight and yield in plants under water deficit in the fruiting stage can be explained by the limitation in water availability and also the local climatic conditions, since according to NÓBREGA et al. (2024) they have high evapotranspiration rates, thus requiring greater availability of water for fruit filling. According to SOARES et al. (2023), the effects of water restriction on plants can be diverse and can affect physiological, growth and biochemical processes, and production. In studies carried out by SIMÕES et al. (2021) with mango, total fruit weight and yield were also more affected by water deficit in the fruiting stage.

For fruit polar diameter (PD), an increase was observed in plants under water deficit in the vegetative/flowering stages and application of ascorbic acid at a concentration of 400 mg L⁻¹ (T19), with an average value of 98.94 mm, an increase of 21.47 mm when compared to those that received 100% ETc during the entire cycle and without application of ascorbic acid (T1).

Foliar application of ascorbic acid promoted a beneficial effect on the polar diameter of guava fruits under water deficit in the vegetative/flowering stage, as well as the relative water content

and electrolyte leakage in the leaf blade; this may be associated with the physiological function of AsA in the detoxification of ROS that are produced under water deficit conditions, as reported by WANG et al. (2018) and NAZ et al. (2016), who observed that, when applied externally, AsA can play a crucial role in the protection of proteins and lipids.

The highest value of soluble solids (SS) was observed in plants subjected to water deficit in the maturation stage and without application of ascorbic acid (T25), which obtained a value of 14.10 °Brix. On the other hand, plants irrigated with 50% ETc and under AsA concentration of 200 mg L⁻¹ in the vegetative stage (T14) obtained the lowest values; when comparing them, an increase of 3.84 °Brix was observed.

Soluble solids contents are significantly influenced by water deficit, as reported by LENG et al. (2024), being an important variable to understand how plants respond to stress conditions. The positive correlation between water deficit and increased levels of sugars such as glucose and fructose may indicate that plants adjust their metabolism to survive under water-restricted conditions (LENG et al., 2024).

In principal component 2, increments in the average fruit weight (AFW) and number of fruits per plant (NF) were observed in plants under full irrigation (100% ETc) and without ascorbic acid application (T1), equal to 203.49 g per fruit and 49.6 units, respectively, while the lowest values were observed in T11 for AFW and T26 for NF. When comparing the highest and lowest values, increments of 69.62 g per fruit and 29.15 units were observed, respectively.

According to ZHONG et al. (2019), there is a tendency for the mass of fruits to increase as water availability increases, which is essential to understand the relationship between irrigation and agricultural yield, corroborating the results obtained in the present study. According to SADIQ et al. (2017), water plays a critical role in several physiological processes in plants, including nutrient uptake, photosynthesis, and fruit formation.

Application of 400 mg L⁻¹ of AsA promoted a beneficial effect on the equatorial diameter of the fruits (ED) of guava cv. Paluma under water deficit in the vegetative/flowering stages (T19), with an increase of 12.01 mm when compared to the fruits of plants subjected to 100% ETc irrigation depth and without application of ascorbic acid (T1).

Equatorial diameter showed the same behavior as polar diameter, and the action of ascorbic acid in improving fruit size may be linked to the fact that AsA contributes to improving several important aspects such as the photosynthetic rate and efficiency

of photosynthesis, thus promoting greater production of carbohydrates and consequently better growth and production of plants (KHAN et al., 2012).

Regarding the vitamin C contents, it was observed that irrigation with 50% ETc in the maturation stage and without application of ascorbic acid (T25) promoted the highest values (47.69 mg 100 g⁻¹ of pulp); when compared to the control treatment T1 (plants irrigated with 100% ETc and without application of ascorbic acid), there was an increase of 9.43%. The increase in vitamin C contents in fruits of guava subjected to water deficit conditions can be explained by its functions, as it is a non-antioxidant compound that increases under water deficit conditions, acting to protect cells against oxidative stress (AKRAM et al., 2017).

Guava pulp pH reached the highest value (4.45) in plants grown under water deficit in the vegetative/flowering stage and foliar application of AsA at a concentration of 400 mL⁻¹ (T19). Conversely, plants grown under full irrigation throughout the cycle and without ascorbic acid application (T1) obtained lower mean values. pH is a crucial factor in evaluating the quality of fruits, including guava. Lower pH values are associated with higher acidity, which can help preserve fruit pulp during storage and processing; in addition, the use of lower pH can allow fruits to be preserved without the need for high heat treatments, which can often result in nutrient loss and change in organoleptic characteristics, such as flavor, color and aroma (BENEVIDES et al., 2008). For titratable acidity (TA), higher mean values and 2.98 % acid citric were found in the fruits of plants subjected to water deficit of 50% ETc in the vegetative/fruiting stages and without application of ascorbic acid (T9), denoting an increase of 22.48% in TTA when compared to the lowest values observed in T1. The increase in titratable acidity in guava fruits is highly desirable in the food industry, as it reduces the need for artificial acidifiers, improving the nutritional and organoleptic quality of the products (BRASIL et al., 2016).

OLIVEIRA et al. (2020) highlighted that natural acidity contributes to the preservation of the sensory attributes of products. SILVA et al. (2018) also reinforced that reducing the use of artificial acidifiers is beneficial to health, improving food quality. Despite the reduction in fruit quality variables in different treatments, the quality of guava pulp meets the quality standard established by Normative No. 49, of September 26, 2018 of the Ministry of Agriculture, Livestock and Food Supply, which defines the minimum parameters for soluble solids (7 °Brix), pH (7.5), titratable acidity (0.4 g

citric acid/100 g pulp) and ascorbic acid (24 mg/100 g pulp) (BRASIL, 2018).

CONCLUSION

Water deficit in the vegetative/fruiting stages negatively affects growth in stem diameter, crown volume, crown diameter and vegetative vigor index, average fruit weight, and polar and equatorial diameter of guava fruits, at 390 days after transplanting. Foliar application of ascorbic acid at a concentration of 600 mg L⁻¹ promotes beneficial effects on total fruit weight and yield of guava. Water deficit in the maturation stage stimulates the synthesis of vitamin C and soluble solids of guava.

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DECLARATION OF CONFLICT OF INTEREST

The authors declare no conflict of interest.

AUTHORS' CONTRIBUTIONS

All authors contributed equally for the conception and writing of the manuscript. All authors critically revised the manuscript and approved of the final version.

DATA AVAILABILITY STATEMENT

The raw data used in this work are available from the first author.

DECLARATION OF USE OF ARTIFICIAL INTELLIGENCE

The authors declare that they did not use artificial intelligence tools to write the manuscript, nor to replace their own skills during its preparation.

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